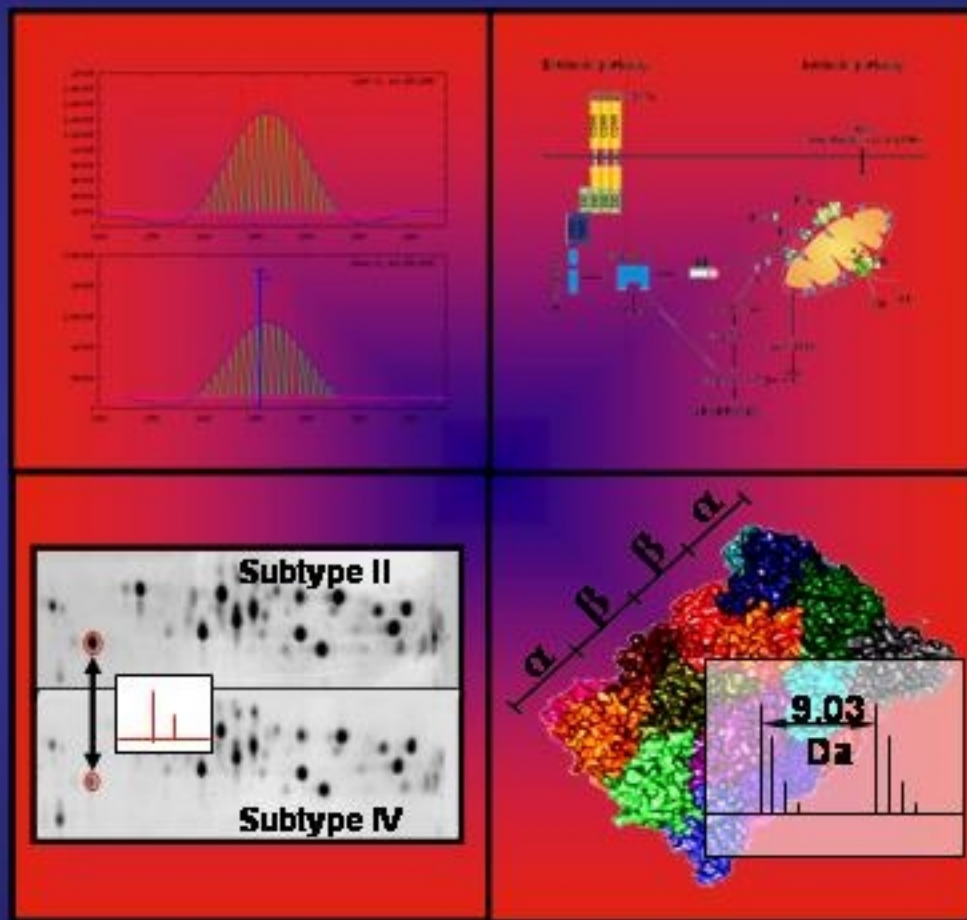




Quantitative proteome analysis of 20S proteasome by ICAT- or SILAC based LC and 2-DE approaches



Dr. Frank Schmidt



Overview:

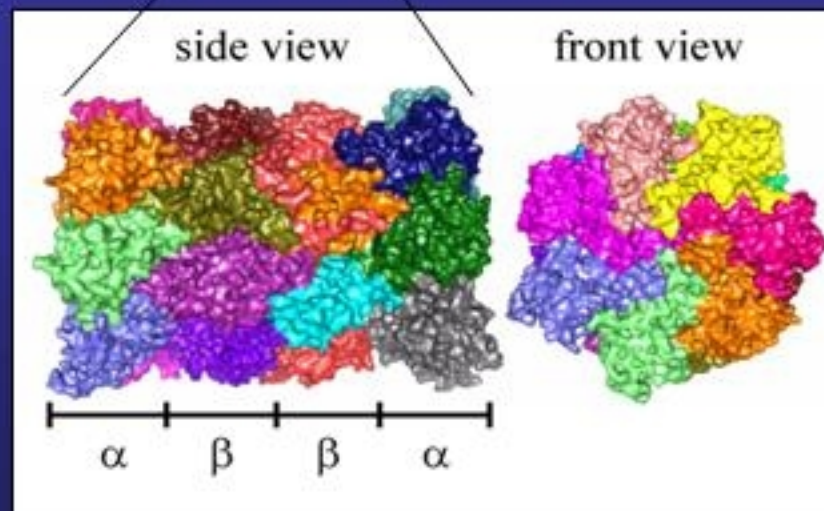
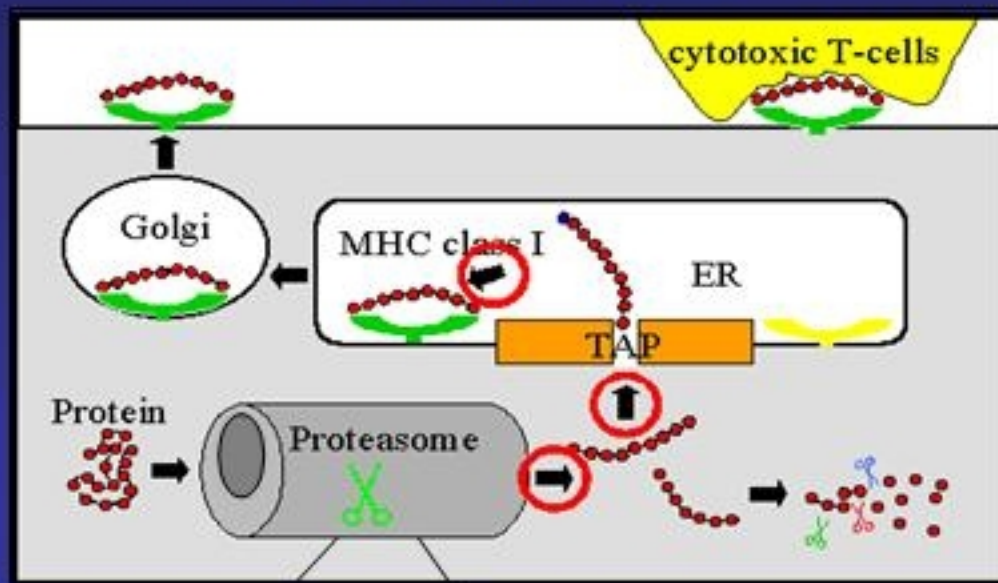
- 1. Quantification of 20S proteasome subtypes by ICAT and 2-DE**
- 2. Quantification of 20S proteasome by SILAC and SDS-PAGE**



Aim of study:

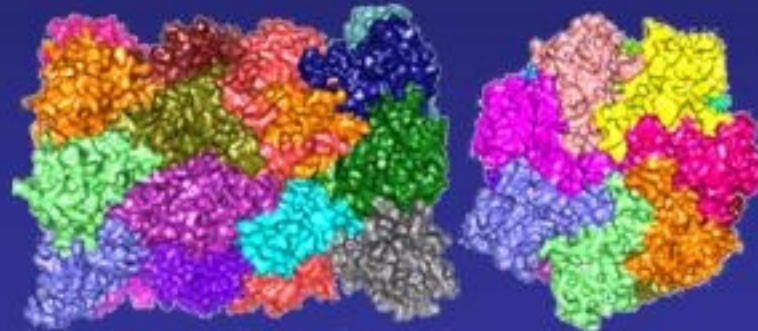
- **Generation of a 2-DE standard pattern of 20S proteasome from rat liver**
- **Find out new PTM's**
- **Comparison of 20S proteasome vs. 20S immunoproteasome using ICAT and 2-DE**
- **Comparison of four common quantification techniques**

1 The Proteasome



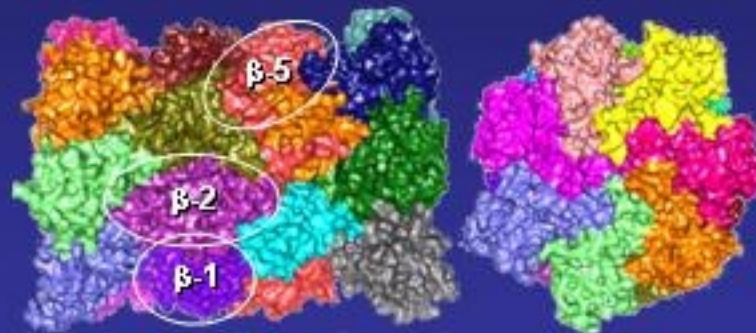
20S Core Complex of the Proteasome

1.1 20S Core Complex and its Cleavage Motifs



- 20S proteasome is composed of 7 α - and 7 β -subunits
- Barrel-shaped heterodimeric configuration: α - β - β - α
- Three nominal activity sites in 20S core are known
- Postglutamyl cleavage activity, catalysed by subunit β -1
- Trypsin-like cleavage, catalysed by subunit β -2
- Chymotrypsin-like cleavage, catalysed by subunit β -5

1.2 Replacement of β 1, 2 and 5 in Immunoproteasome



- 3 of 7 β -subunits were replaced in the immunoproteasome
- Transformation of β -1 into β -1i, β -2 into β -2i and β -5 into β -5i
- β -i forms are different in its AA composition and leads to a new conformation and change in the proteasome netto charge
- Changes are detectable by ion-exchange chromatography

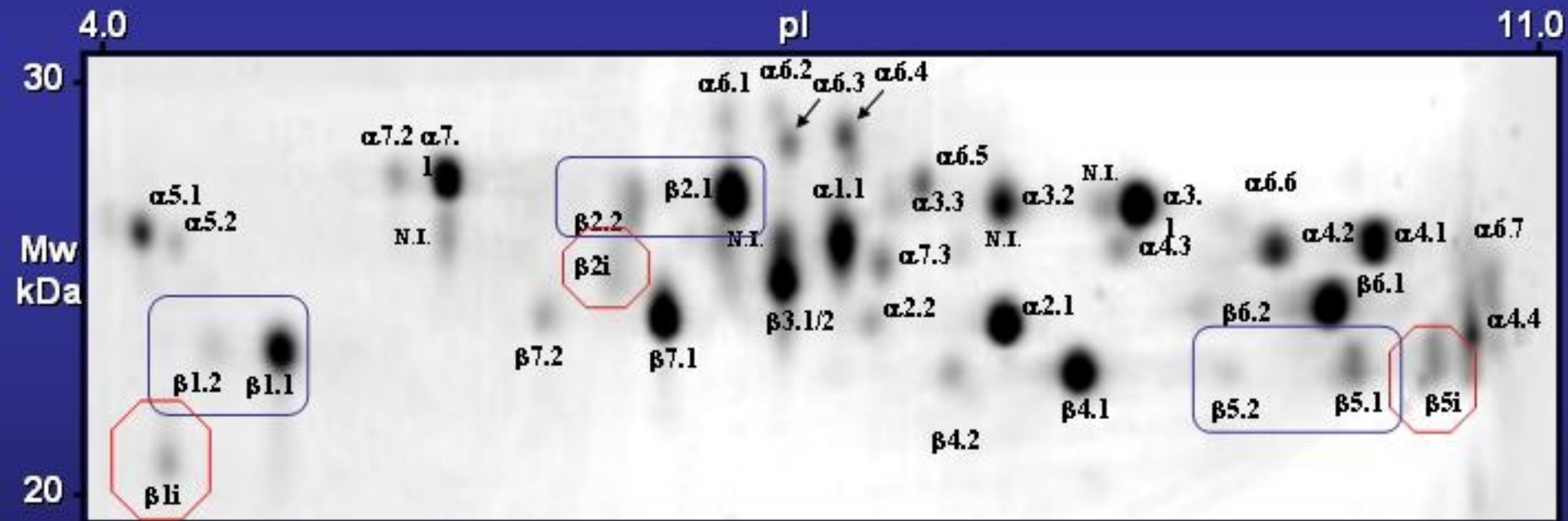


1.3 2DE Standard Pattern of the 20S Core Complex

Gel-chromatography

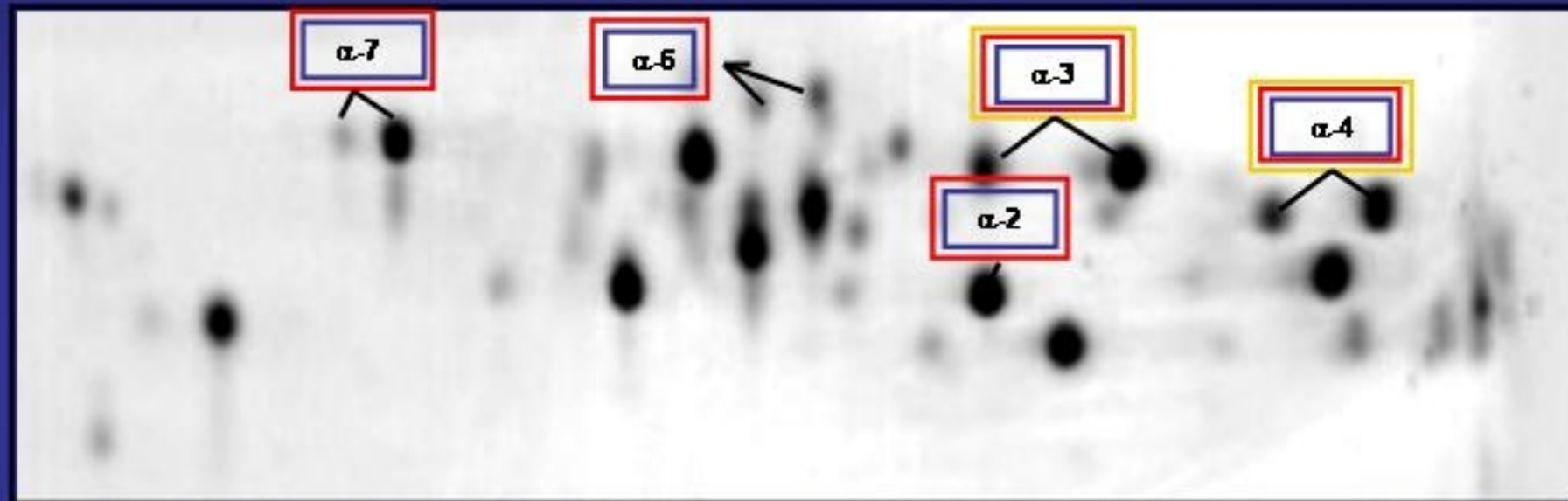
Ion-exchange chromatography

72 protein species separated by 2-DE

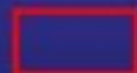




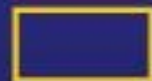
1.4 Known Post-translational Modifications



Phosphorylation



N-Acetylation

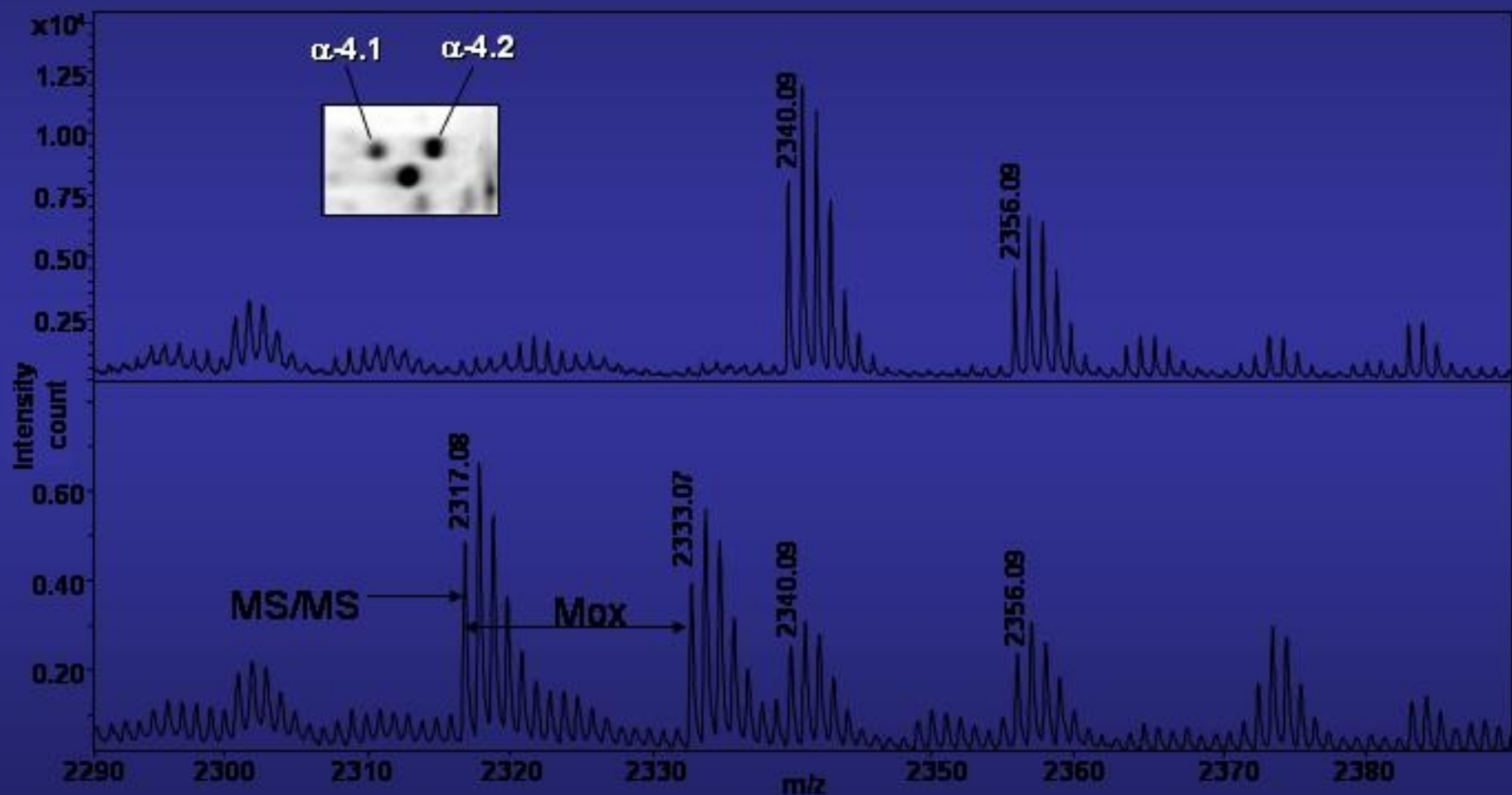


Glycosylation

Deamidation: α -7

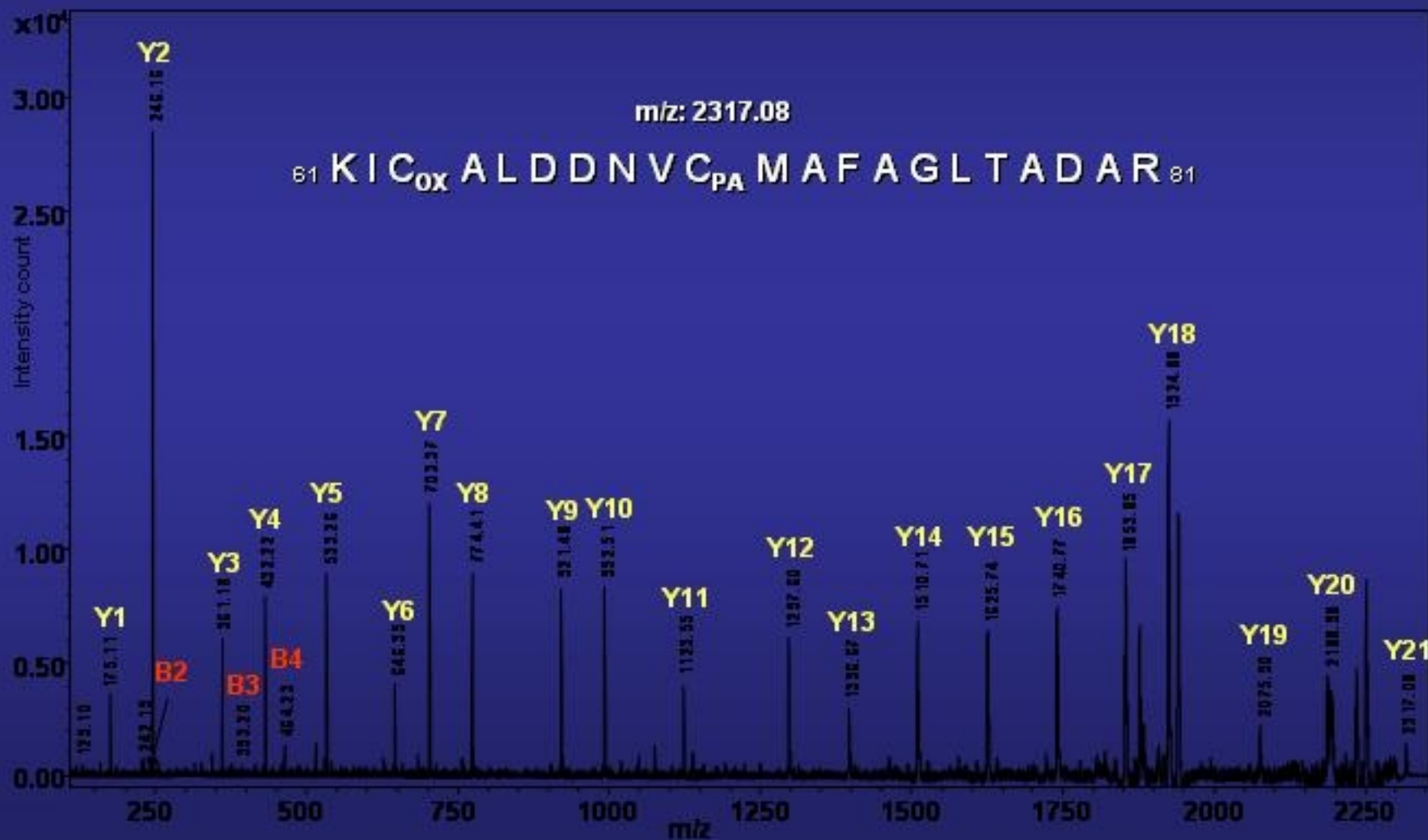


1.5 New PTM: Cysteine Oxidation at Subunit α - 4





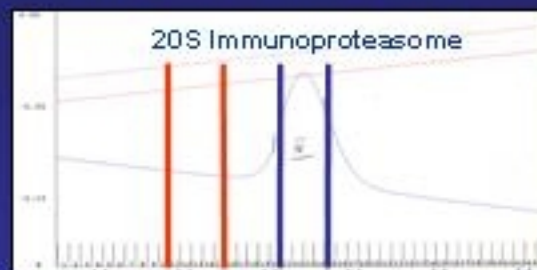
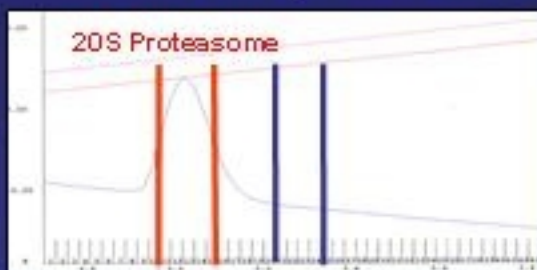
1.6 New PTM: Cysteine Oxidation at Subunit α – 4





1.8 Analytical Course of Action: Quantification

Ion-exchange chromatography



Labelling
cICAT-¹²C

Labelling
cICAT-¹³C

Combination

LC/ESI-MS/MS

LC/MALDI-MS/MS

2-DE Gel
MALDI-MS

2-DE Gel
MALDI-MS

SEQUEST
Xpress

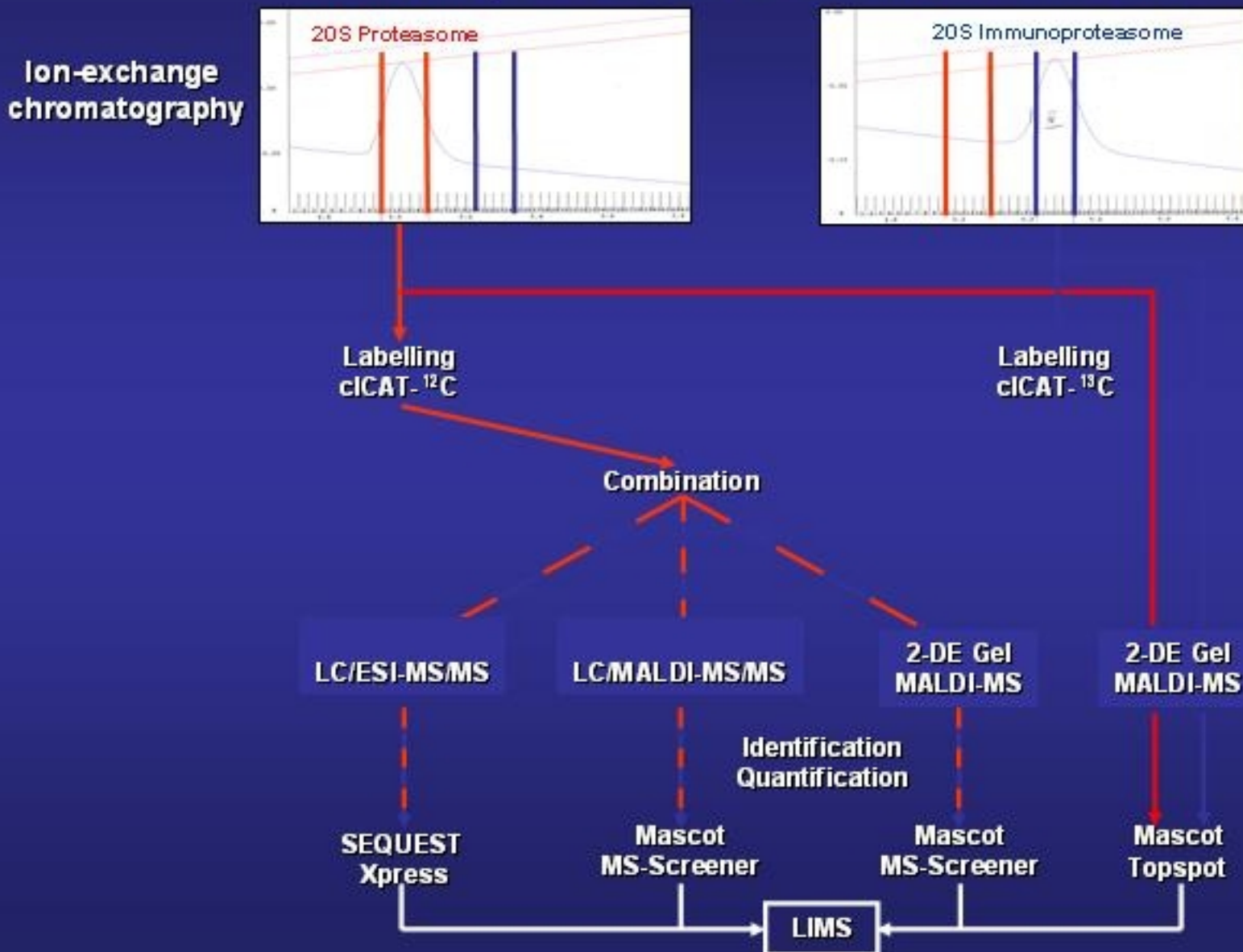
Mascot
MS-Screener

Identification
Quantification

Mascot
MS-Screener

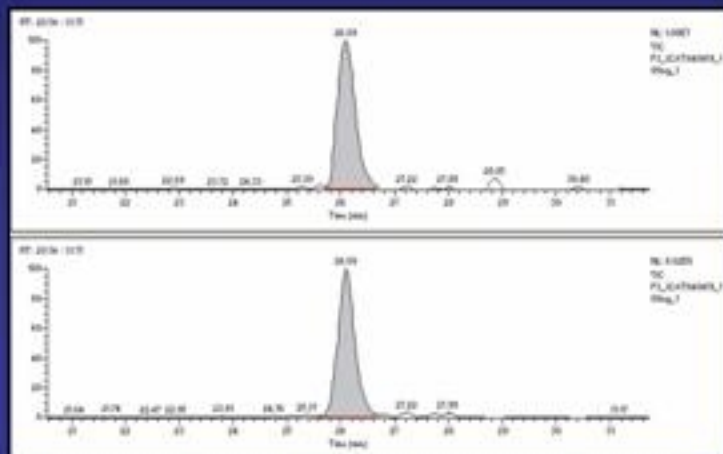
Mascot
Topspot

LIMS

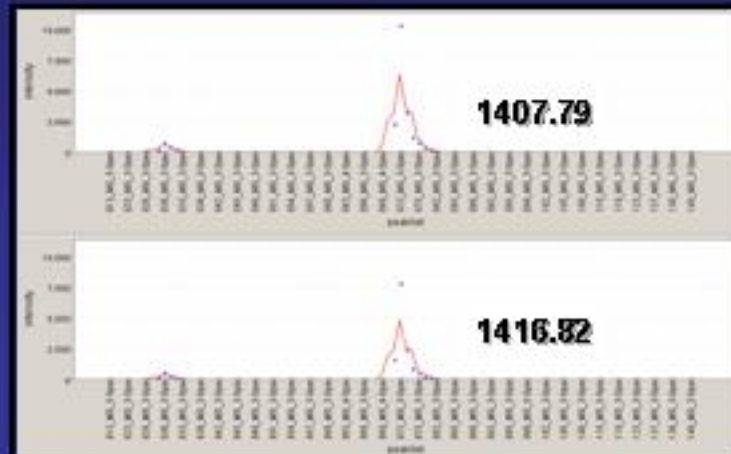




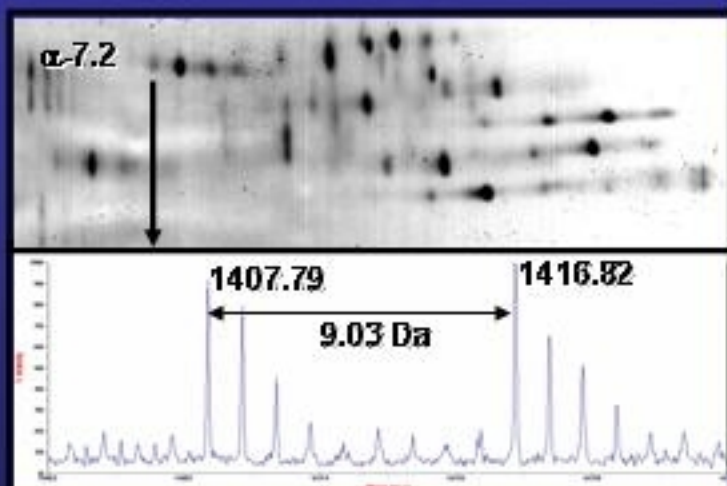
1.9 Quantification Techniques



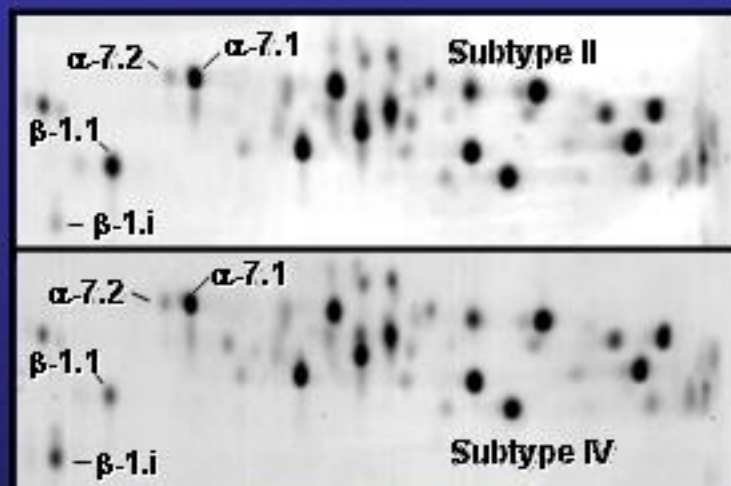
cICAT/LC/ESI-MS



cICAT/LC/MALDI-MS



cICAT/2-DE/MALDI-MS



2-DE/MALDI-MS

1.10 Results Quantification Techniques

| Criteria/Methods | ICAT based | | | |
|---------------------------------|------------|----------|-------------|-------------|
| | LC-ESI | LC-MALDI | 2-DE/ MALDI | 2-DE/ MALDI |
| Time effort | ~5h | ~20h | ~14d | ~14d |
| Sensitivity | 200ng | 200ng | 30µg | 30µg |
| Protein coverage | 70% | 53% | 100% | 100% |
| Quantitative standard deviation | +/- 20% | +/- 5% | n.a. | +/- 20% |
| Sequence coverage proteins | 10-20% | 10-20% | 30-70% | 30-70% |
| PTM's | N | N | 56 | 56 |



Conclusion

- Development of an 2-DE standard pattern of 20S rat liver proteasome
- Identification of all protein species only by 2-DE
- PTM's were only detected by 2-DE (56)
- New cysteine oxidation at α -4.1 discovered
- ICAT and 2-DE are complementary approaches
- 2-DE spots with more than one protein species can only be quantified at peptide level (isotopic labelling)

- Replacement of β -subunits 1 were detected by all approaches
- Replacement of β -subunits 2 and 5 were only detected by 2-DE
- ICAT/LC-MALDI was the most accurate method (+/- 5% StDv)

Schmidt et al. (2006) *Proteomics* 3: 24-32.

Schmidt et al. (2003) *Mol Cell Proteomics* 3: 24-42.

Schmidt et al. (2003) *J Am Soc Mass Spectrom* 14: 943-956.



2. Quantification of 20S Proteasome by SILAC and SDS-PAGE

Aim of study:

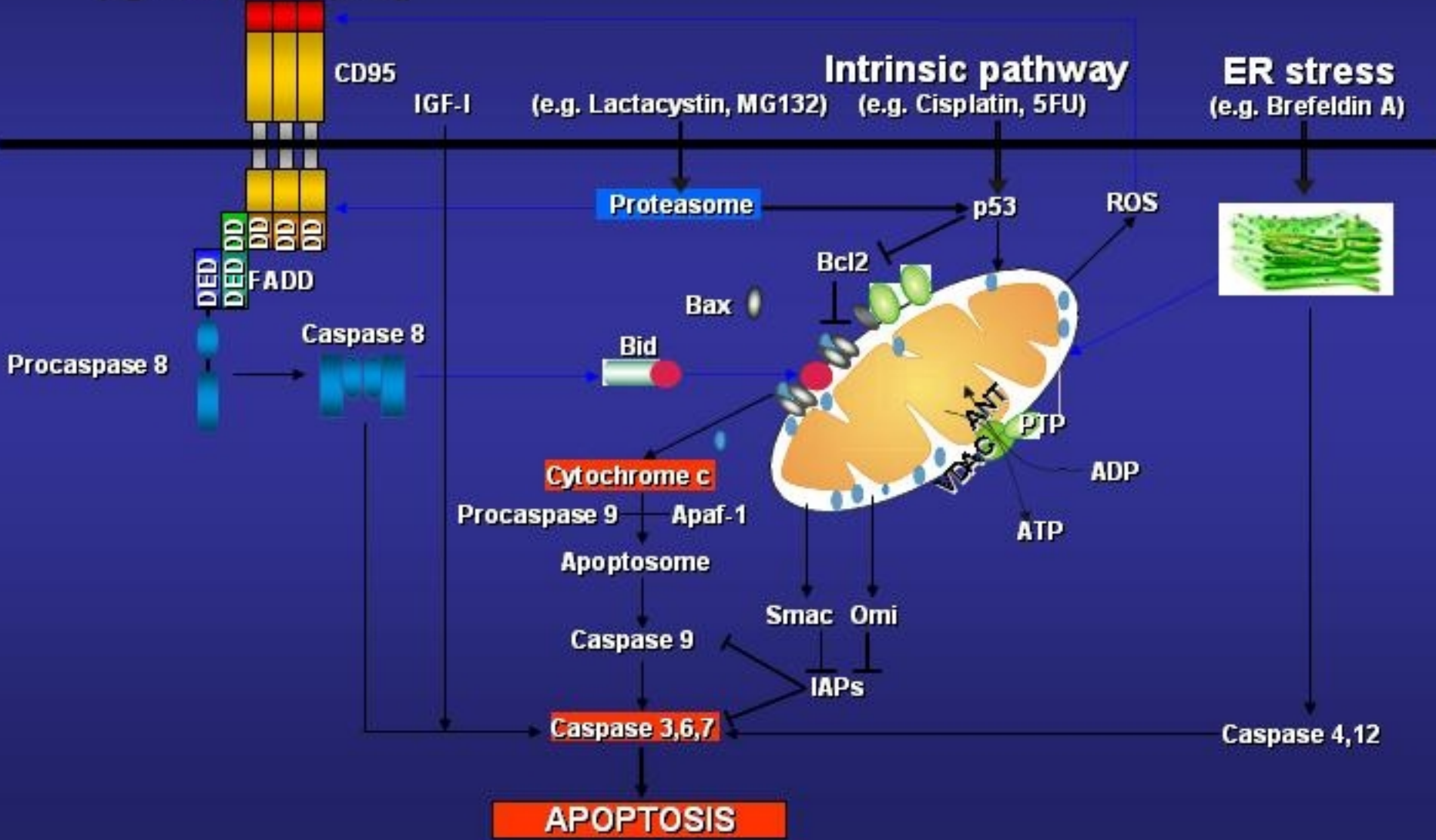
**-Quantification of 20S proteasome from apoptosis induced vs. healthy cells
by SDS-PAGE and LC-MS**



2.1 Apoptosis Signalling

Extrinsic pathway (e.g. CD95L (FasL/Apo-1L))

(e.g. CD95L (FasL/Apo-1L))



APOPTOSIS



2.2 20S Proteasome Quantification of 5FU Induced Apoptosis

Stable Isotope Labelling by Amino Acids in Cell Culture (SILAC)

$^{12}\text{C}_6$ Arginine/ Lysine (light)



$^{13}\text{C}_6$ Arginine/ Lysine (heavy)



Jurkat T-Cells

"Health" vs. Cisplatin

Gel-chromatography



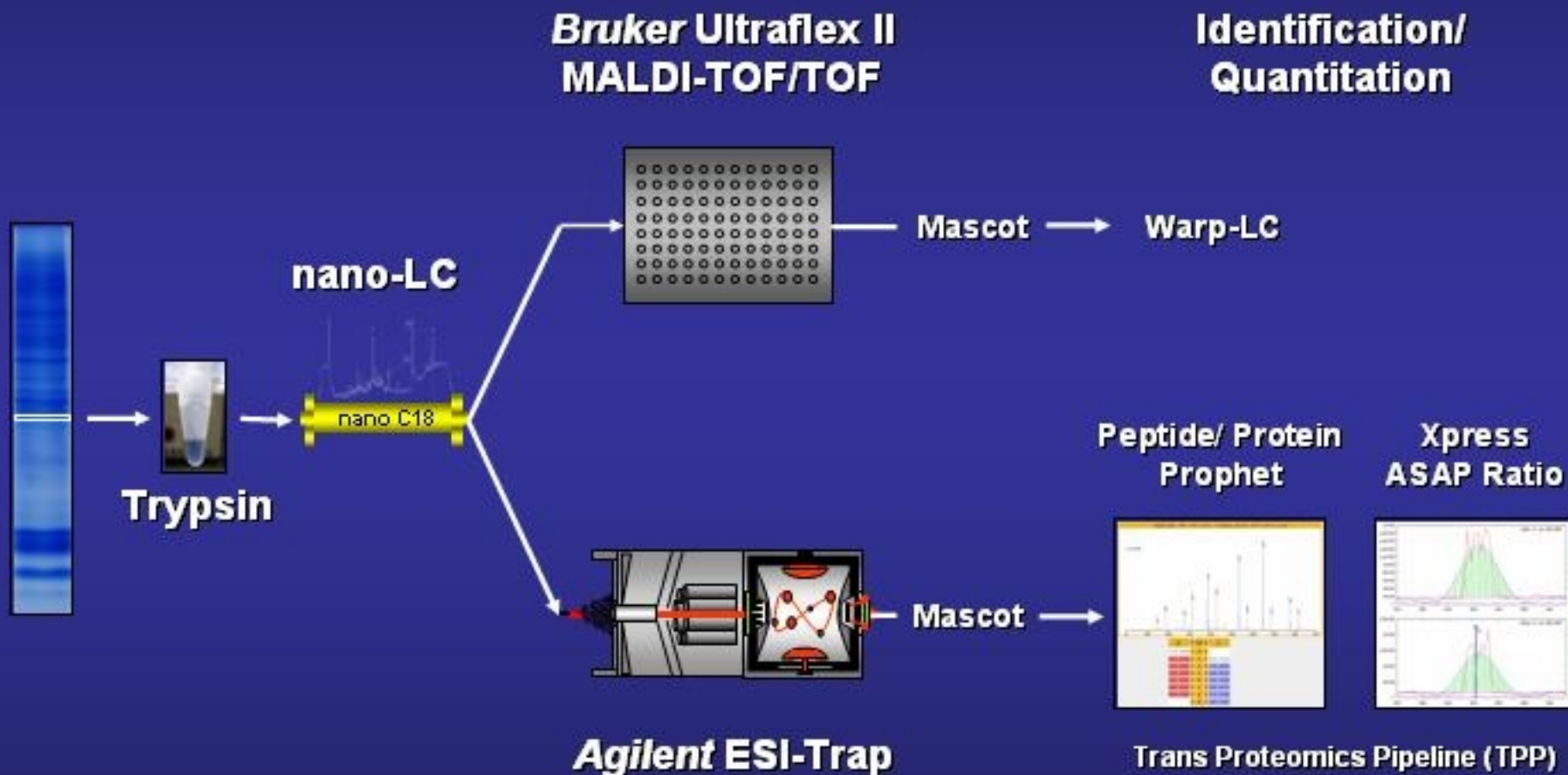
Protein separation



SDS-PAGE



2.3 Experimental Course of Action





2.4 Quantification Results LC-MALDI-MS Health vs. Apoptotic

| Subunit | Acc. # | Quantified Peptides | Ratio H/L | StDv |
|-------------|--------|---------------------|-------------|------|
| PSA1 | P25786 | 6 | 1,00 | 0,07 |
| PSA2 | P25787 | 6 | 1,16 | 0,12 |
| PSA3 | P25788 | 4 | 0,64 | 0,06 |
| PSA4 | P25789 | 5 | 1,00 | 0,09 |
| PSA5 | P28066 | 4 | 1,03 | 0,06 |
| PSA6 | P60900 | 4 | 1,00 | 0,01 |
| PSA7 | O14818 | 6 | 1,01 | 0,06 |
| PSB1 | P20618 | 3 | 1,06 | 0,10 |
| PSB2 | P49721 | 4 | 1,05 | 0,06 |
| PSB3 | P49720 | 4 | 1,03 | 0,17 |
| PSB5 | P28074 | 7 | 0,99 | 0,16 |
| PSB6 | P28074 | 3 | 0,99 | 0,16 |
| PSB7 | Q99436 | 2 | 0,98 | 0,01 |



2.5 Quantification Results LC-ESI-MS Health vs. Apoptotic

| Subunit | Acc. # | Quantified Peptides | Ratio H/L | StDv |
|---------|---------|---------------------|-----------|------|
| PSA1 | P25786 | 6 | 0.79 | 0,22 |
| PSA2 | P25787 | 8 | 1.00 | 0,24 |
| PSA4 | P25789 | 2 | 1.00 | 0,02 |
| PSA5 | P28066 | 6 | 0.97 | 0,15 |
| PSA6 | P60900 | 9 | 0.83 | 0,32 |
| PSA7 | O14818 | 15 | 0.92 | 0,25 |
| PSB1 | P20618 | 10 | 1.09 | 0,14 |
| PSB2 | P49721 | 6 | 0.96 | 0,38 |
| PSB3 | P49720 | 9 | 1.04 | 0,29 |
| PSB4 | P28070 | 10 | 0.92 | 0,31 |
| PSB5 | P28066 | 15 | 0.87 | 0,36 |
| PSB6 | P280721 | 8 | 1.15 | 0,24 |
| PSB7 | Q994361 | 4 | 1.24 | 0,26 |
| PSB8 | P280621 | 5 | 1.05 | 0,18 |
| PSB9 | P280651 | 6 | 0.95 | 0,34 |
| PSB10 | P403061 | 1 | 0.48 | 0,06 |



2.4 Conclusion

LC-MALDI-MS

- 13 of 17 subunits detected
- Average StDv +/- 9%

LC-ESI-MS

- 16 of 17 subunits detected
- Average StDv +/- 23%

- Decreased amount of PSA3 in apoptotic cells
- Immunoproteasome subunits PSB1i,2i and 5i only by LC-ESI-MS
- LC-ESI showed a higher number of lysine containing peptides
- LC-MALDI showed higher number for arginine containing peptides
- Protein species not distinguishable (Quantification: sum of protein species)
- low number of PTM's
- LC-MALDI showed the most accurate quantification



Acknowledgement

Biotechnology Centre Oslo, Norway

Dr. B. Thiede, H. Hustoft, M. Strozinsky

Max Planck Institute Berlin, Germany

Dr. P. Jungblut, M. Schmid, R. Ackermann

Charité Berlin, Germany

Prof. B. Dahlmann, Dr. K. Janek, A. Kloss